

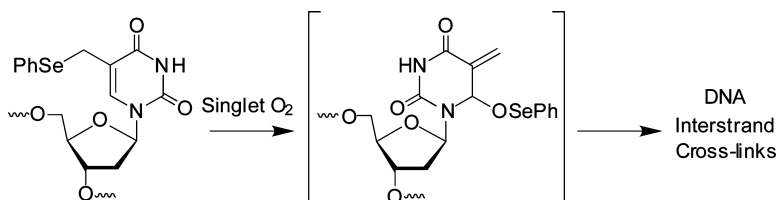
Communication

**DNA Interstrand Cross-Link Formation Initiated by Reaction
 between Singlet Oxygen and a Modified Nucleotide**

In Seok Hong, and Marc M. Greenberg

J. Am. Chem. Soc., **2005**, 127 (30), 10510-10511 • DOI: 10.1021/ja053493m • Publication Date (Web): 07 July 2005

Downloaded from <http://pubs.acs.org> on March 25, 2009



More About This Article

Additional resources and features associated with this article are available within the HTML version:

- Supporting Information
- Links to the 10 articles that cite this article, as of the time of this article download
- Access to high resolution figures
- Links to articles and content related to this article
- Copyright permission to reproduce figures and/or text from this article

[View the Full Text HTML](#)

DNA Interstrand Cross-Link Formation Initiated by Reaction between Singlet Oxygen and a Modified Nucleotide

In Seok Hong and Marc M. Greenberg*

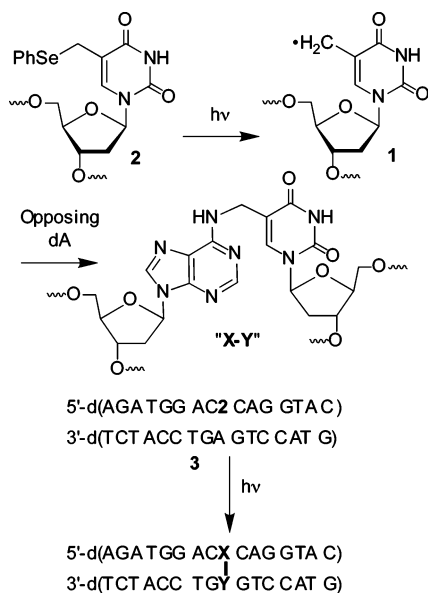
Department of Chemistry, Johns Hopkins University, 3400 North Charles Street, Baltimore, Maryland 21218

Received May 28, 2005; E-mail: mgreenberg@jhu.edu

DNA is often the target of anti-cancer agents, which alkylate or oxidatively damage the biopolymer. Hydroxyl radical, metal-oxo complexes, such as that produced by bleomycin, and singlet oxygen are examples of agents that oxidatively damage DNA.^{1–4} Singlet oxygen, which selectively oxidizes deoxyguanosine, is an important reactive oxygen species in photodynamic therapy.^{3,5,6} The reactivity of a subset of DNA alkylating agents is distinguished from reactive oxygen species by their formation of interstrand cross-links (ISC's). For instance, interstrand cross-links are believed to be the source of the cytotoxicity of the anti-cancer agents, mitomycin C and chlorambucil.⁷ Herein, we describe a process involving a modified nucleotide (**2**) that potentiates the effects of singlet oxygen by reacting with this reagent to form ISC's.

We recently described a mechanism in which 5-(2'-deoxyuridinyl)methyl radical (**1**) forms an ISC with the opposing deoxyadenosine when it is photochemically generated from **2** in DNA (Scheme 1).^{8,9} In the course of investigating the mechanism for

Scheme 1



this process, we examined the effect of singlet oxygen on DNA containing **2**. Filtered ($\lambda \geq 400$ nm) aerobic photolysis (30 min) of 5'-³²P-**3** in the presence of 1–50 μ M of the singlet oxygen sensitizer, Rose Bengal, produced ISC's in as high as 48% yield (Figure 1B). Anoxic photolysis of a mixture of 5'-³²P-**3** and Rose Bengal (50 μ M) produces ~3% ISC.¹⁶ These observations suggest that direct photolysis of the phenyl selenide (**2**), which generates ISC's via **1** independent of O₂, and in lower yield, is not the source of cross-links under these conditions. Furthermore, the anoxic results suggest that ISC's do not result from a direct photoreaction between the sensitizer and DNA. Instead, the dependence of the rate of

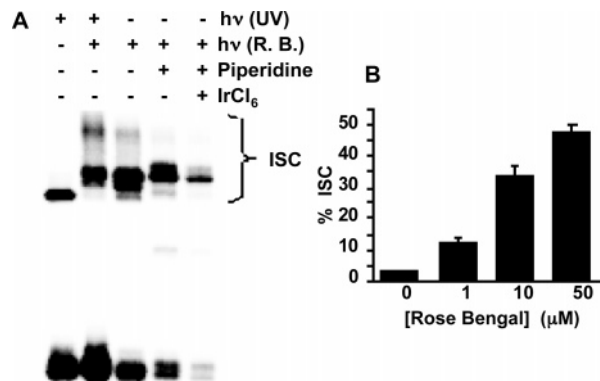


Figure 1. Formation of DNA interstrand cross-links (**3**, 10 nM) via UV or Rose Bengal sensitized aerobic photolysis. (A) Autoradiogram comparing ISC's produced upon UV or Rose Bengal (50 μ M) sensitized photolysis. (B) Effect of Rose Bengal concentration on ISC formation (30 min photolysis).

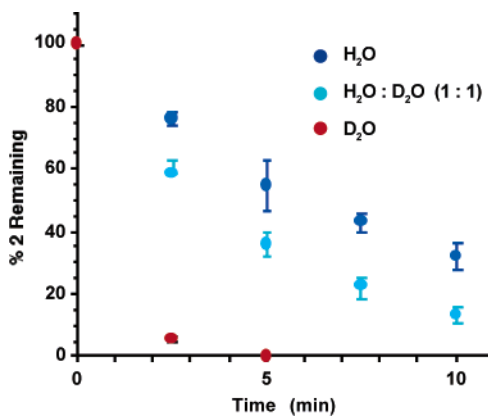
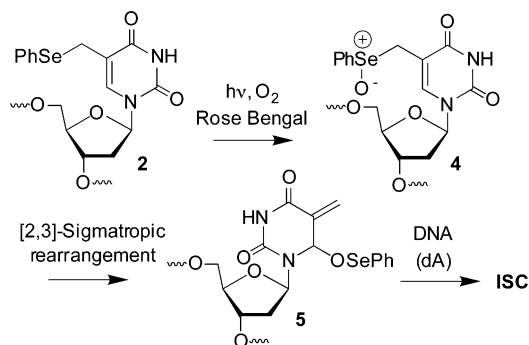


Figure 2. Effect of D₂O on the consumption of monomeric **2** (50 μ M) upon irradiation of Rose Bengal (10 μ M).

disappearance of monomeric **2** on D₂O content suggests that singlet oxygen, whose lifetime is enhanced 10-fold in the deuterated solvent, is responsible for phenyl selenide consumption (Figure 2).¹⁰

Photolysis of an otherwise identical duplex containing dT in place of **2** produces ~2% ISC, indicating that the phenyl selenide (**2**) plays an integral role in their formation in **3**.¹⁶ In contemplating a mechanism for this process, we recognized that phenyl selenides are oxidized to selenoxides by singlet oxygen in good yield, and allylic selenoxides undergo [2,3] sigmatropic rearrangements.^{11,12} Execution of these reactions in **2** would produce an electrophilic methide-type intermediate (**5**), akin to other molecules that alkylate DNA (Scheme 2).^{13,14} Evidence for this mechanism was gleaned from NMR analysis of the reaction of monomeric **2** with NaIO₄ (Figure 3) or its photosensitization by Rose Bengal in deuterated phosphate buffer.¹⁶ Periodate is shown because it rapidly and completely oxidizes **2**. The phenyl selenide (**2**) is completely

Scheme 2



consumed within 10 min and replaced by a diastereomeric mixture of the rearrangement product (**5**). Selenoxide **4** is not detected. Compound **5** reacts with the weak nucleophilic H₂O over the course of 24 h to produce 5-hydroxymethyl-2'-deoxyuridine (**6**), which is identical to independently prepared material.¹⁵ Furthermore, ISC's are efficiently formed with the opposing 2'-deoxyadenosine upon treatment of **3** with NaIO₄, indicating that the methide (**5**) produces cross-links.¹⁶

When **5** is produced in duplex DNA, rotation about the N-glycosidic bond into the *syn*-conformation positions the exocyclic methylene to react with N1 of the opposing deoxyadenosine, which is the same position that **1** is believed to cross-link with. However, the cross-link products formed in the presence of singlet oxygen migrate more slowly than those produced via **1** (Figure 1A). Exposure of 5'-³²P-**3** to singlet oxygen (Figure 1A, lane 2) previously cross-linked via formation of **1** (Figure 1A, lane 1) indicates that the cross-links produced under singlet oxygen conditions contain additional damage. Oxidized deoxyguanosines

within the ISC products were deemed to be the most likely lesions formed in addition to cross-links. The damaged purines were revealed by treatment of the ISC's with piperidine and IrCl₆ (Figure 1A, lanes 4 and 5).^{17–20} Piperidine treatment following oxidation with IrCl₆ converts many of the ISC products into shorter, faster migrating fragments, indicating that not all of the cross-linked products also contain additional damaged nucleotides.¹⁶

The reaction of **2** with singlet oxygen is also distinguished from the radical pathway (**1**) by the effect of glutathione on ISC formation. ISC formation is unaffected by physiologically relevant glutathione concentrations (5 mM) when **3** is subjected to singlet oxygen.¹⁶ This observation reinforces those reported above, which suggest that the combination of the phenyl selenide derivative of thymidine (**2**) and singlet oxygen offers a novel and potentially important means for producing interstrand cross-links in DNA. The physiological importance of interstrand cross-links suggests that the incorporation of phenyl selenide **2** in DNA could be an effective adjuvant in photodynamic therapy.

Acknowledgment. We are grateful for support from the National Institute of General Medical Sciences (GM-054996).

Supporting Information Available: Experimental procedures for carrying out the experiments on **2** and **3**. Autoradiograms showing ISC formation from **3** by NaIO₄ and hydroxyl radical footprinting of these ISC's, effect of glutathione on ISC formation, decomposition of singlet oxygen induced ISC's by piperidine treatment, and control experiments. ¹H NMR showing formation of **5** by Rose Bengal sensitized photolysis of **2**. This material is available free of charge via the Internet at <http://pubs.acs.org>.

References

- Chen, J.; Stubbe, J. *Nat. Rev. Cancer* **2005**, *5*, 102–112.
- von Sonntag, C. *The Chemical Basis of Radiation Biology*; Taylor & Francis: London, 1987.
- Lee, P. C. C.; Rodgers, M. A. J. *Photochem. Photobiol.* **1987**, *45*, 79–86.
- McCallum, J. E. B.; Kaniyoshi, C. Y.; Foote, C. S. *J. Am. Chem. Soc.* **2004**, *126*, 16777–16782.
- Dolmans, D. E.; Fukumura, D.; Jain, R. *Nat. Rev. Cancer* **2003**, *3*, 380–387.
- Prat, F.; Hou, C.-C.; Foote, C. S. *J. Am. Chem. Soc.* **1997**, *119*, 5051–5052.
- Schärer, O. D. *ChemBioChem* **2005**, *6*, 27–32.
- Hong, I. S.; Greenberg, M. M. *J. Am. Chem. Soc.* **2005**, *127*, 3692–3693.
- Monomeric compounds are referred to by the same number as when in **3**.
- Turro, N. J. *Modern Molecular Photochemistry*; Benjamin Cummings: Menlo Park, CA, 1978.
- Krief, A.; Lonzé, F. *Tetrahedron Lett.* **2002**, *43*, 6255–6257.
- Reich, H. J.; Yelm, K. E.; Wollowitz, S. *J. Am. Chem. Soc.* **1983**, *105*, 2503–2504.
- Douki, T.; Vadesne-Bauer, G.; Cadet, J. *J. Org. Chem.* **2003**, *68*, 478–482.
- Veldhuyzen, W. F.; Shallop, A. J.; Jones, R. A.; Rokita, S. E. *J. Am. Chem. Soc.* **2001**, *123*, 11126–11132.
- Sowers, L. C.; Beardsley, G. P. *J. Org. Chem.* **1993**, *58*, 1664–1665.
- See Supporting Information.
- Muller, J. G.; Duarte, V.; Hickerson, R. P.; Burrows, C. J. *Nucleic Acids Res.* **1998**, *26*, 2247–2249.
- Ye, Y.; Muller, J. G.; Luo, W.; Mayne, C. L.; Shallop, A. J.; Jones, R. A.; Burrows, C. J. *J. Am. Chem. Soc.* **2003**, *125*, 13926–13927.
- Duarte, V. G. D.; Yamaguchi, L. F.; Ravanat, J. L.; Martinez, G. R.; Medeiros, M. H. G.; DiMascio, P. D.; Cadet, J. *J. Am. Chem. Soc.* **2000**, *122*, 12622–12628.
- Ravanat, J.-L.; Di Mascio, P.; Martinez, G. R.; Medeiros, M. H. G.; Cadet, J. *J. Biol. Chem.* **2000**, *275*, 40601–40604.

JA053493M

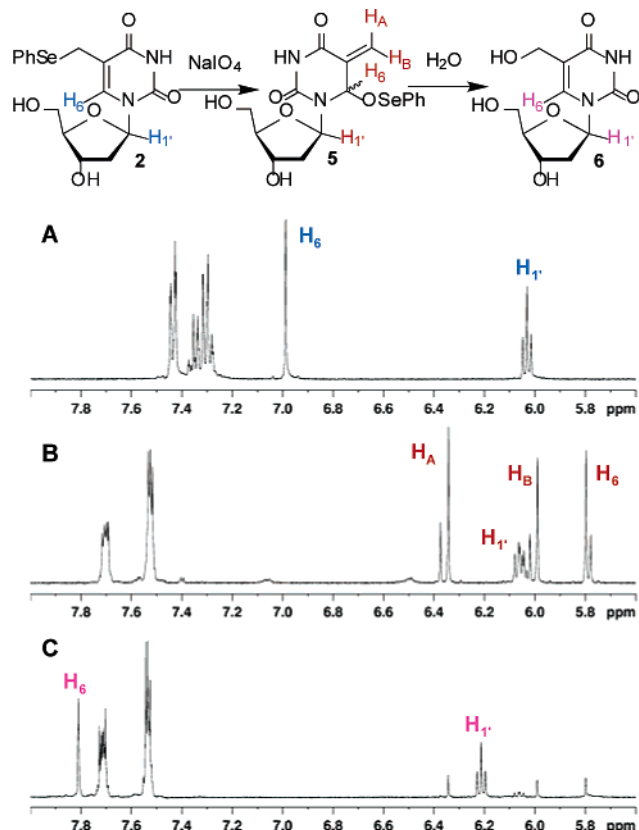


Figure 3. ¹H NMR analysis of the reaction of **2** (50 mM) with NaIO₄ (50 mM) in deuterated phosphate buffer (50 mM, pD 7.4). (A) Before NaIO₄ addition, (B) 10 min after NaIO₄ addition, and (C) 24 h after NaIO₄ addition.